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(54) Title: **PCR-HYBRIDIZATION ASSAYS SPECIFIC FOR INTEGRATED RETROVIRUSES**

(57) Abstract: The present invention provides a method of determining latent HIV infection by detection low-abundant HIV nucleic acid sequences in samples of cellular DNA or in paraffin-fixed tissue sections or frozen sections. This technique calls for one primer for an Alu sequence in the human chromosomal DNA and the other primer near the end of the viral LTR. PCR-amplified DNA is then hybridized with a probe which only recognizes the amplified LTR sequence. This probe could be labeled in different ways, and quantitative assays for latent HIV or other retoviruses infection could be developed.

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PCR-HYBRIDIZATION ASSAYS SPECIFIC FOR INTEGRATED RETROVIRUSES

5

10

BACKGROUND OF THE INVENTION

Field of the Invention

The present invention relates generally to the fields of
15 virology and diagnostics. More specifically, the present invention
relates to the development of PCR-hybridization assays specific
for integrated retroviruses and their use in the determination of
latent infection in patients.

20 Description of the Related Art

It has been known for many years that latent HIV
infection can occur in infected cell lines *in vitro* and nearly all of
the early studies of the molecular mechanisms underlying HIV
latency examined what is occurring in latently infected clonally
25 derived T-cell lines (2-4, 10, 13, 15). It was also demonstrated
many years ago that there is a considerably greater frequency
(~100-fold more) of lymphocytes in HIV patients which contain
HIV DNA than cells which are productively infected (i. e., possess
HIV DNA as well as making HIV RNA, viral proteins, and progeny

virions) (6, 7). Studies examining HIV infection of resting or activated CD4 lymphocytes *in vitro* clearly showed that only activated cells are primarily permissive for virus replication and allow progeny virus production, and infection of resting cells usually leads an abortive infection, in which the virus binds, enters, and completely or partially reverse transcribes proviral DNA. This viral DNA, however, does not integrate until the cell is activated into division within a few days (9, 17). Most of the relatively higher frequency of HIV DNA⁺/RNA⁻ lymphocytes in HIV patients (~0.8%) (5), therefore, are very likely recently infected (abortively) resting lymphocytes. This should be expected since 99% of all lymphocytes are resting (16). However, some of these HIV DNA⁺ /RNA⁻ cells found in infected subjects may also be latently infected cells, and these cells would have integrated proviral DNA but are not making progeny virus. The frequency of latently infected cells in HIV⁺ individuals is not clear.

Studies *in vitro* examining acute and chronic infection of both human T-cell lines and PHA-stimulated CD4 PBLs has shown that gradual shutdown of the virus into latency is routine for the majority of acutely infected cells, and shutdown into latency should be considered a normal part of the virus life cycle (11). Thus, in an HIV patient, any productively infected cells, which are not eliminated by the immune system or by replication of cytopathic HIVs, should become latently infected cells. Thus, it is possible that reliable quantitative data on the frequencies of latently infected cells in patients may reveal that there exists a large pool of such cells.

One study attempted to directly quantitate the frequency of latently infected cells in HIV patients using an

inefficient re-ligation technique of PCR-amplified HIV-cellular DNA and found that the frequency was low: at approximately the same frequency as productively-infected cells (~ 7 cells per 10^6) (5). More recent studies have used limiting dilution culturing of lymphocytes from patients, which are activated by PHA to reactivate latent virus. The frequencies of these cells are much higher (8). Unfortunately, this procedure also allows any recently abortively infected resting cells to complete the virus replication cycle and become HIV-producing cells, and thus those studies do not discriminate completely between the abortively infected cells and latently infected cells.

Understanding the extent of latent infection has become of great importance now because HAART therapy can reduce plasma viral loads for long periods of time, yet when the drugs are removed, virus usually returns. It seems reasonable to conjecture that a great deal of this return is due to reactivation of HIV from latently infected cells.

The prior art is deficient in the lack of effective means of detecting latent HIV infection in patients. The present invention fulfills this long-standing need and desire in the art.

SUMMARY OF THE INVENTION

The present invention describes an *in situ* PCR assay that only detects integrated retroviruses, and this should allow accurate assessment of the frequency of productively and latently infected cells together in patients. Since the frequencies of productively infected cells at any given time in patients is known

to be very low, any larger frequencies would have to be measuring latent infection. Through the use of PCR and hybridization, a new way to score for integrated retroviruses in cellular chromosomal DNA is possible. This method may be adapted in many forms of quantitative assays of latent HIV infection in infected subjects undergoing anti-retroviral drug therapy and should allow physicians to monitor the presence of latently infected cells so that they can determine whether treatment should be discontinued or not.

The invention utilizes PCR and employs one primer for an Alu sequence in the human chromosomal DNA and the other primer near the end of the viral LTR. PCR-amplified DNA is then hybridized with a probe which only recognizes the amplified LTR sequence. The probe could be labeled in different ways, and numerous forms of this test could be developed, many of which could be quantitative.

In one aspect of the present invention, there is provided a quantitative PCR assay to measure HIV latent infection.

In another aspect of the present invention, there is provided a kit for detecting integrated retroviruses in a human sample.

In another embodiment, the invention can be used to monitor latent infection by other retroviruses.

Other and further aspects, features, and advantages of the present invention will be apparent from the following description of the presently preferred embodiments of the invention given for the purpose of disclosure.

BRIEF DESCRIPTION OF THE DRAWINGS

So that the matter in which the above-recited features, advantages and objects of the invention, as well as
5 others which will become clear, are attained and can be understood in detail, more particular descriptions of the invention briefly summarized above may be had by reference to certain embodiments thereof which are illustrated in the
10 appended drawings. These drawings form a part of the specification. It is to be noted, however, that the appended drawings illustrate preferred embodiments of the invention and therefore are not to be considered limiting in their scope.

Figure 1 shows primer design and probe hybridization. One primer lies in an *alu* sequence in the human
15 chromosomal DNA and the other primer is located near the end of the viral LTR. PCR amplified DNA is then hybridized with a probe which only recognizes the amplified LTR sequence.

Figure 2A shows results from *in situ* PCR-hybridization on uninfected CEM cells, productively-infected 35.8
20 cells, a fifty-fifty mixture of the two, and on HIV latently infected CEM cells. No reaction occurred in the uninfected cells, but strong reactions occurred in both the productively infected and latently infected cells.

Figure 2B shows HIV p24 immunostaining of CEM cell
25 populations used in *in situ* PCR hybridization assays. Uninfected CEM cells (negative control); productively-infected 35.8 cells (positive control); latently-infected CEM cells show less than 5% of these cells are making HIV.

Figure 2C shows *in situ* PCR-hybridization on CEM cells and latently-infected CEM with single primers alone. Performing the assay with only the alu primers or the LTR primers alone did not give any reaction.

5 Figure 3A shows *in situ* PCR-hybridization reactions on uninfected, acutely-infected, and latently-infected purified CD4 PHA-blasts. Cells were fixed and paraffin-embedded, and sections were made.

10 Figure 3B shows HIV p24 immunostaining of uninfected, acutely-infected, and latently-infected purified CD4 PHA-blasts. The latently-infected PBLs are negative.

15 Figure 4 shows acute abortive infection is negative by this assay. Resting CD4 lymphocytes were acutely infected and 18 hours later were cytocentrifuged onto glass slides and *in situ* PCR-hybridization was performed.

20 Figure 5 shows standard DNA PCR on DNA extracted from the abortively infected resting cells in Figure 4 using HIV gag primers. This shows that HIV DNA is present in these cells but it is in the unintegrated form. DNA extracted from uninfected cells serve as a control.

DETAILED DESCRIPTION OF THE INVENTION

25 Assessment of whether cells possess integrated retrovirus in their chromosomal DNA is relatively straightforward when all the cells examined have at least one viral copy. This can be accomplished by routine Southern blot hybridization techniques of either uncut or restriction enzyme digested

chromosomal DNA. However, in the case of HIV infection, much less than 1% of cells contain integrated retroviruses, and PCR techniques have to be used.

Currently, clinicians can only monitor newly produced
5 virus by the quantitative RT-PCR assays or the presence of HIV
DNA by PCR (99% of which represents abortive infection of newly
infected cells and not cells making virus). There is no easy way
to monitor the extent of latent infection. The present invention is
a new way to score for integrated retroviruses in cellular
10 chromosomal DNA, which could be exploited for quantitative
assays of productive plus latent HIV infection in patients
undergoing anti-retroviral drug therapy. This technique utilizes
PCR for the detection of low-abundant HIV nucleic acid sequences
only in cellular DNA and is described for *in situ* assays on
15 paraffin-fixed tissue sections or frozen sections, although it could
be used in any number of formats. The invention features one
primer for an Alu sequence in the human chromosomal DNA and
the other primer near the end of the viral LTR (see Fig. 1). PCR-
amplified DNA is then hybridized with a probe which only
20 recognizes the amplified LTR sequence. Numerous forms of this
test could be developed, many of which could be quantitative.

The DNA hybridization probe can be labelled by any
of the many different methods known to those skilled in this art.
The labels most commonly employed for these studies are
25 radioactive elements, enzymes, chemicals which fluoresce when
exposed to ultraviolet light, and others. A number of fluorescent
materials are known and can be utilized as labels. These include,
for example, fluorescein, rhodamine, auramine, Texas Red, AMCA
Blue and Lucifer Yellow. The radioactive label can be detected by

any of the currently available counting procedures. The preferred isotope may be selected from ^3H , ^{14}C , ^{32}P , ^{35}S , ^{36}Cl , ^{51}Cr , ^{57}Co , ^{58}Co , ^{59}Fe , ^{90}Y , ^{125}I , ^{131}I , and ^{186}Re .

Enzyme labels are likewise useful, and can be detected
5 by any of the presently utilized colorimetric, spectrophotometric, fluorospectrophotometric, amperometric or gasometric techniques. The enzyme is conjugated to the selected particle by reaction with bridging molecules such as carbodiimides, diisocyanates, glutaraldehyde and the like. Many enzymes which
10 can be used in these procedures are known and can be utilized. Those of ordinary skill in the art will know of suitable labels which may be employed in accordance with the present invention.

In one embodiment of the present invention, there is
15 provided a quantitative method of detecting integrated retroviruses in a human sample by first amplifying DNA in the sample with PCR using one primer for an Alu sequence and the other primer for a retroviral sequence. Preferably, the primer for the Alu sequence has the sequence of 5'-
20 TCCCAGCTACTGGGGAGGTGAGG-3' (SEQ ID NO: 1), and the primer for the retroviral sequence recognizing the 5' LTR sequence of HIV has the sequence of 5'-AAGCTCGATATCAGCAGTTCT-3' (SEQ ID NO: 2). Alternatively, a primer for the 3' LTR (such as 5'-CAGCTGCTTTTTGCCTGTACTG-3', SEQ ID NO: 3) could be used.
25 The PCR-amplified DNA is then hybridized with a probe that specifically recognizes the amplified retroviral sequence. Preferably, the samples are paraffin-embedded sections or frozen sections of blood, lymph nodes or lymphocytes. In another embodiment of the invention, this technique can be used to

monitor latent infection by other retroviruses through the use of primers that specifically recognize the LTR sequences of other retroviruses.

In another aspect of the present invention, there is
5 provided a kit for detecting integrated retroviruses and/or
determining the number of cells with integrated retroviruses in a
human sample. The kit contains a primer for an Alu sequence, a
primer for a retroviral sequence, a probe that specifically
recognizes amplified retroviral sequence, and a detection agent
10 that detects hybridization of the probe to amplified retroviral
sequence. Preferably, the primer for the Alu sequence has the
sequence 5'-GCCTCCCAAAGTGCTGGGATTACAG-3' (SEQ ID NO: 4),
and the primer for the retroviral sequence recognizing the 5' LTR
sequence of HIV has the sequence of 5'-
15 AAGCTCGATATCAGCAGTTCT-3' (SEQ ID NO: 2). The kit can be
used on samples of paraffin-embedded sections or frozen
sections of blood, lymph nodes or lymphocytes. In another
embodiment of the invention, the kit can be used to detect other
integrated retroviruses and/or determining the number of cells
20 with other integrated retroviruses by using primers that
specifically recognize the LTR sequences of other retroviruses.

The following examples are given for the purpose of
illustrating various embodiments of the invention and are not
meant to limit the present invention in any fashion.

25

EXAMPLE 1

Cell culture

Peripheral blood lymphocytes isolated from human
5 blood by centrifugation through LSM were PHA-stimulated and
grown in RPMI-1640 medium, supplemented with 15% FBS and 40
U/ml of IL-2. Uninfected cells served as negative controls and
cultures infected with laboratory isolates of HIV (AC1 and R28)
served as positive control samples. Also, CEM cells served as
10 negative controls, while a clonal CEM line constitutively
producing HIV (35.8 cells) (11) which harbor one to two copies
of integrated HIV provirus served as positive control.

15

EXAMPLE 2

In situ PCR and hybridization

3 x 10⁵ cells in 50 µl of PBS were cytocentrifuged at
1000 rpm for 2 min onto glass slides. The slides were then
20 dipped in 4% paraformaldehyde (PFA) at RT for 20 min. They
were then transferred to 3X PBS (stop reaction) at RT for 2 min.,
to 1X PBS for 2 min., and transferred to fresh 1X PBS for 2 min.
The slides were then dehydrated through 50%, 70% 100% ethanol
for 4 min/each and allowed to air dry. The slides were heated at
25 100°C at a heating block for 90 sec. and then treated with 0.3%
H₂O₂ at RT overnight. The next day the slides were washed and
immersed in diluted proteinase K in PBS (6µg/ml) at RT for 5 min.
The slides were heated at 95°C for 2 min. to stop the reaction,
rinsed in PBS for 10 min. and rinsed in H₂O for 10 min. They

were then air-dried. 50 μ l of amplification cocktail (30.5 μ l dH₂O, 5.0 μ l 10X PCR buffer, 4.0 μ l dNTP, 1.0 μ l primer1, 1.0 μ l primer2, 8.0 μ l MgCl₂, 0.5 μ l Taq) was added into each well, and a coverslip was carefully sealed around the edge. The slides were heated on

5 a heating block at 92° C for 90 sec. PCR was performed at 95°C for 40 sec., 55°C for 90 sec., and 72°C for 90 sec. for 30 cycles. The slides were soaked in 100% ethanol and heated at 92°C for 1 min. The slides were soaked in 2X SSC at RT for 5 min., and placed at 4°C overnight. 100 μ l of hybridization mix (50 μ l

10 deionized Formamide, 10 μ l 20X SSC, 10 μ l 100X Denhart's sol, 10 μ l ssDNA (10mg/ml), 10 μ l 10% SDS, 4 μ l dH₂O, and 6 μ l probe (25-50mg/ml)) was added into each well on the slide and heated at 95°C for 5 min. Then slides were placed on ice for 1min. and incubated at 42°C for 2-4 hours with coverslips in a humid

15 chamber. The slides were washed in 2X SSC at RT: 5min x 2 times, and 0.1X SSC at 42°C for 10min. The sections were dipped into buffer 1 (0.1M tris-HCl, 0.15M NaCl, pH 7.5) and covered with 20-40 μ l buffer 2 (0.5% blocking reagent in buffer 1). Coverslips were added, and the slides were placed at RT for 15

20 min. The coverslips were removed and the slides were dipped in buffer 1. Twenty μ l of diluted anti-Digoxigenin Ab 1:500 in buffer 2 was added to each slide, coverslipped, and incubated at RT in humid chamber for 1 hr. The coverslips were removed and the slides were washed with buffer 1: 10min x 2 times, and

25 equilibrated for 5 min in buffer 3 (color detection B). 20 μ l NBT/BCIP was added onto each section, and coverslips were replaced. The slides were incubated overnight in the dark and were observed the next day under a microscope.

EXAMPLE 3

Latent infection in cloned or normal T cells

Uninfected, latently-infected and productively-
5 infected CEM T-cells were cytocentrifuged onto glass slides, fixed
in paraformaldehyde, and the *in situ* PCR assay was performed as
described. Figure 2 shows that no reaction occurred in the
uninfected cells (CEM), but strong blue reactions occurred in
both of the chronically, productively-infected (35.8), as well as
10 the latently infected cells. Figure 2B shows which cells above
were producing HIV by *in situ* fluorescent immunostaining for
HIV p24. Performing the PCR with only the Alu primer or the LTR
primer alone did not give any reaction (Fig. 2C), and, therefore,
this PCR assay appeared to be specific for the presence of HIV
15 DNA. The majority of latently-infected cells were not making HIV
P24 by *in situ* immunostaining (Fig. 2B).

In situ PCR-hybridization reactions were also
performed on uninfected and acutely-infected purified CD4 PHA-
blasts. These cells were fixed, paraffin-embedded, and sectioned.
20 Strong blue reactions occurred in infected lymphocytes but not in
the uninfected cells (Fig. 3). Again, latently-infected PBLs were
positive but P24-negative (Fig. 3B). Thus, the technique works
well on paraffin-embedded normal lymphocytes.

To show definitively that this procedure only detects
25 integrated HIV DNA, resting CD4 lymphocytes were acutely
infected and 18 hrs later were cytocentrifuged onto glass slides,
and the *in situ* PCR-hybridization was performed (Fig. 4). The
abortively-infected resting cells were negative, but DNA extracted
from the cells was HIV-DNA positive by routine PCR (Fig. 5).

Thus, this *in situ* PCR-hybridization technique only detects integrated, not unintegrated, HIV DNA,

The following references were cited herein:

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25

Any patents or publications mentioned in this specification are indicative of the levels of those skilled in the art to which the invention pertains. These patents and publications are herein incorporated by reference to the same extent as if

each individual publication was specifically incorporated by reference.

One skilled in the art will readily appreciate that the present invention is well adapted to carry out the objects and
5 obtain the ends and advantages mentioned, as well as those inherent therein. The present examples along with the methods, procedures, treatments, molecules, and specific compounds described herein are presently representative of preferred
10 embodiments, are exemplary, and are not intended as limitations on the scope of the invention. Changes therein and other uses will occur to those skilled in the art which are encompassed within the spirit of the invention as defined by the scope of the claims.

WHAT IS CLAIMED IS:

1. A method of detecting integrated retroviruses in a human tissue sample, comprising the steps of:

5 amplifying DNA in the sample with PCR using one primer for an Alu sequence and the other primer for a retroviral sequence;

 hybridizing the PCR-amplified DNA with a probe that specifically recognizes the amplified retroviral sequence; and

10 detecting hybridization of the probe, wherein the hybridization indicates the presence of integrated retroviruses in the sample.

15 2. The method of claim 1, wherein the tissue sample is selected from the group consisting of paraffin-embedded tissue sections and frozen tissue sections.

20 3. The method of claim 1, wherein the tissue sample is selected from the group consisting of lymphocytes, blood, and lymph nodes.

25 4. The method of claim 1, wherein the primer for the retroviral sequence recognizes the LTR sequence of HIV.

5. The method of claim 4, wherein the primer has the sequence of SEQ ID NO: 2.

5 6. The method of claim 4, wherein the primer has the sequence of SEQ ID NO: 3.

7. The method of claim 1, wherein the primer for
10 the Alu sequence has the sequence of SEQ ID NO: 1.

8. The method of claim 1, wherein the method quantitatively determines the number of cells with integrated HIV
15 in the human tissue sample.

9. A kit for detecting integrated retroviruses and/or determining the number of cells with integrated
20 retroviruses in a human tissue sample, said kit containing:

- a primer for an Alu sequence;
- a primer for a retroviral sequence;
- a probe that specifically recognizes amplified retroviral sequence; and

25 a detection agent that detects hybridization of the probe to the amplified retroviral sequence.

10. The kit of claim 9, wherein the primer for the Alu sequence has the sequence of SEQ ID NO: 4.

5 11. The kit of claim 9, wherein the primer for the retroviral sequence recognizes the LTR sequence of HIV.

12. The kit of claim 11, wherein the primer has the
10 sequence of SEQ ID NO: 2.

13. The kit of claim 9, wherein the human tissue sample is selected from the group consisting of paraffin-
15 embedded tissue sections and frozen tissue sections.

14. The kit of claim 9, wherein the human tissue sample is selected from the group consisting of lymphocytes,
20 blood, and lymph nodes.

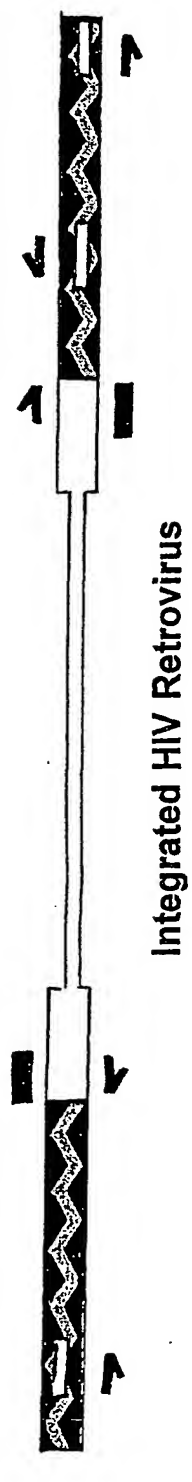


Fig. 1

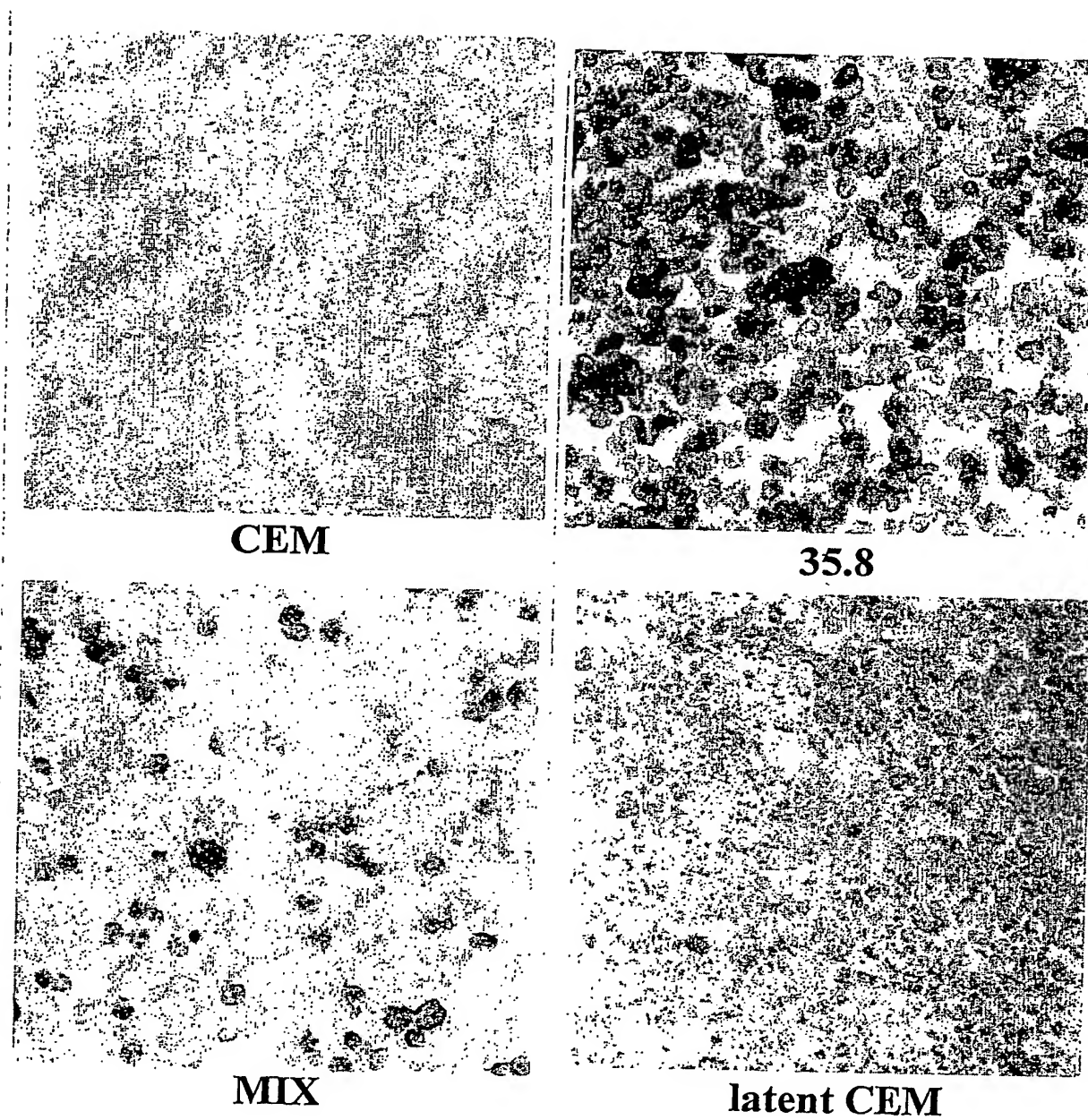
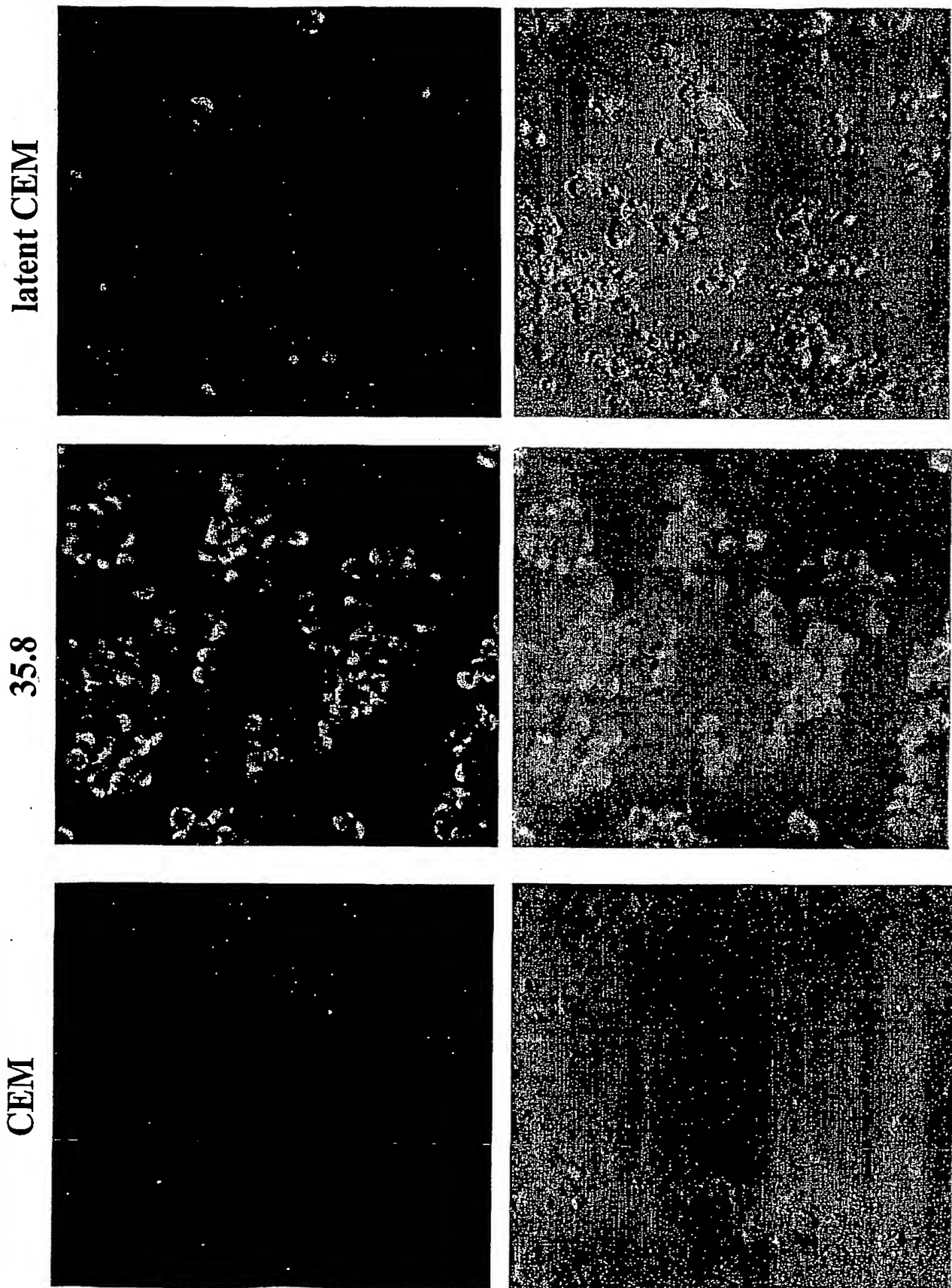
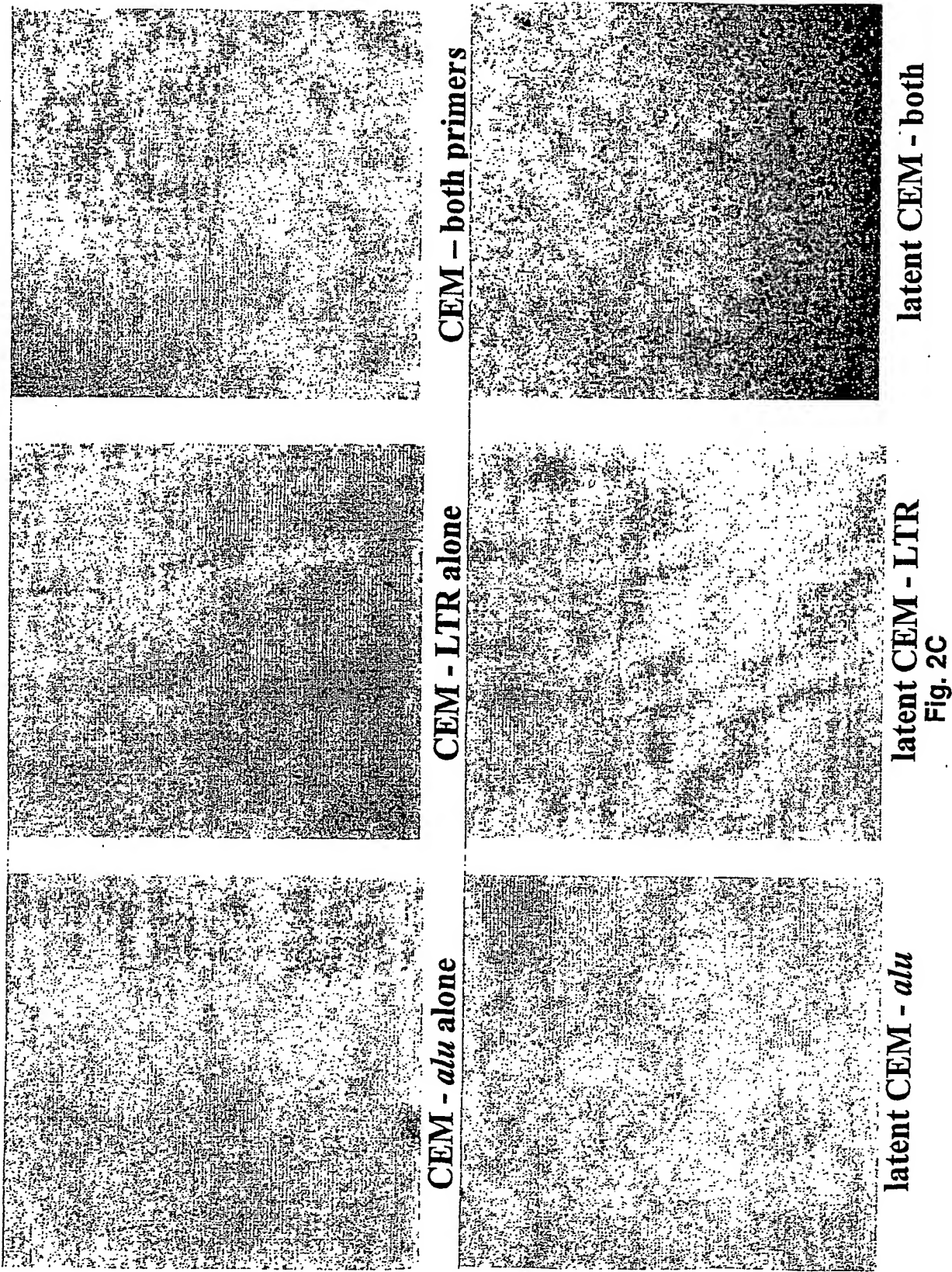


Fig. 2A





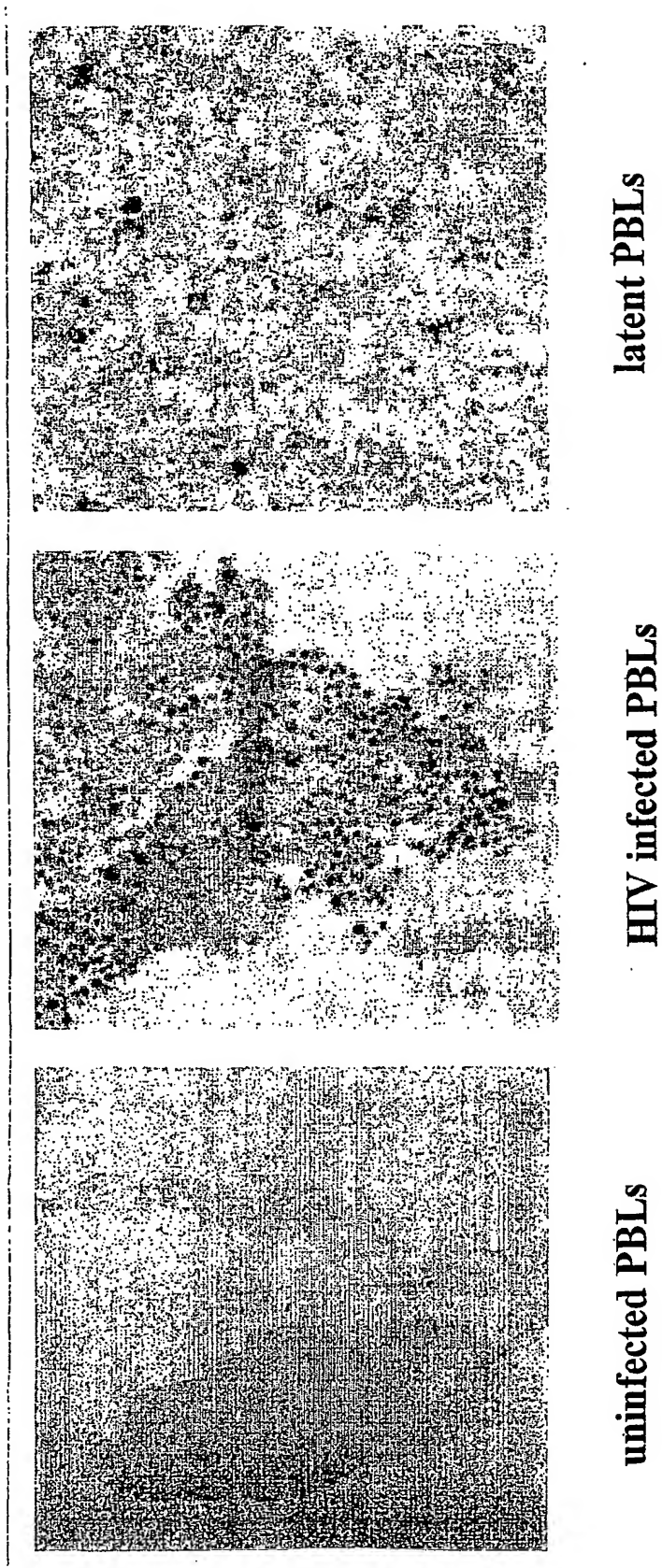
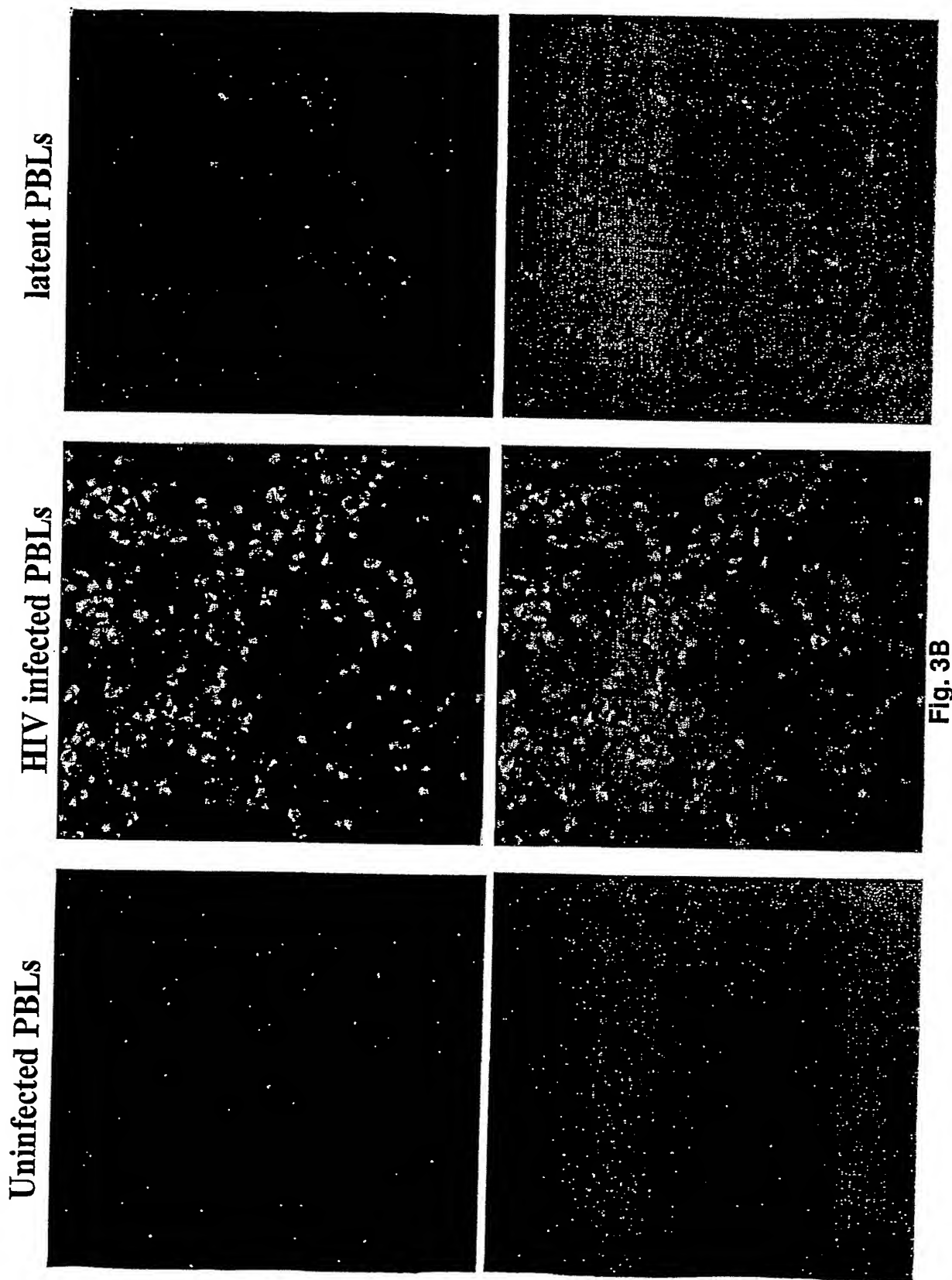
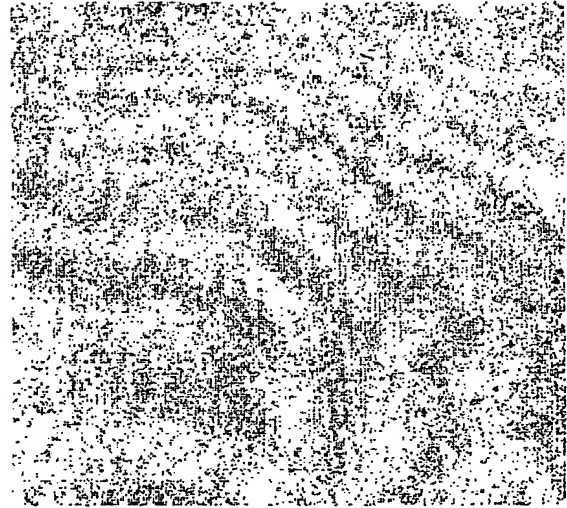


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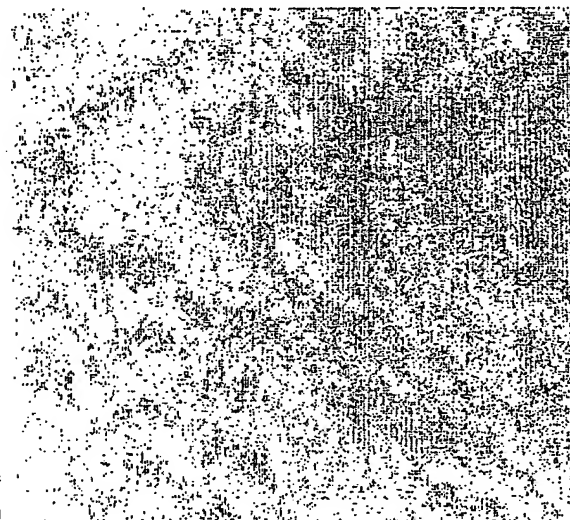




uninfected, resting PBLs



HIV infected, resting PBLs



uninfected, PHA-stimulated PBLs



HIV infected, PHA-stimulated PBLs

Fig. 4

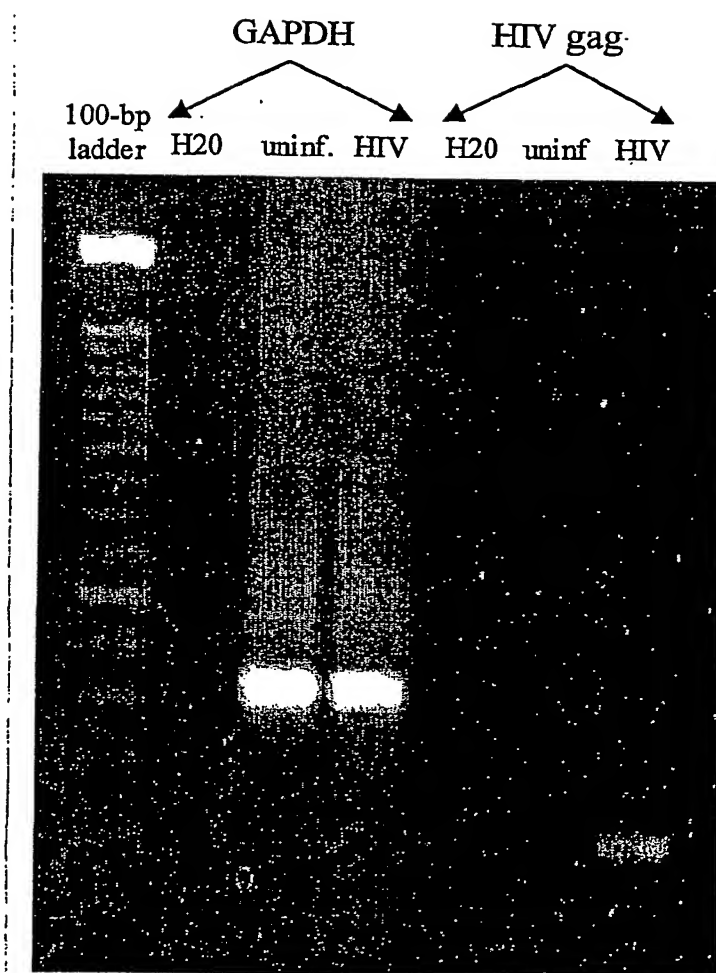


Fig. 5

SEQUENCE LISTING

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